Comparison of stress genes in switchgrass (*Panicum virgatum* L.) genotypes under drought conditions

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Switchgrass (*Panicum virgatum* L.) is being examined as a viable source for biomass feedstock. It is a native, warm-season perennial that belongs to the grass family (Poaceae). Because it is able to grow on marginal lands with minimum to no fertilizer requirements, and few obvious pest and disease issues, this plant could have great success as a novel agronomically produced crop. Since water is required for plants to grow, any change in water availability can have a significant impact on plant growth and development.

Because of this, breeding of more drought-tolerant cultivars is an on-going endeavor. Genes being expressed under such abiotic stresses have been identified in model organisms as well as agriculturally important crops. In this study we measured the expression of known drought-related genes; *rd22*, *rab18*, and *dreb2a*. The three genes were compared between the two different switchgrass genotypes; Alamo and Cave-In-Rock. The plants were grown under normal water conditions (control) and drought induced conditions (treated), in order to analyze the variation in expression.

### 1. Introduction

Drought response is a multigenic trait; the increase of a single gene’s mRNA expression is not a direct correlation for tolerance, but can be used for a better understanding of the switchgrass germplasm for future purposes such as traditional breeding or gene transformation. Perform more biological replicates of the qPCR procedure. Cultivate and test different genotypes in the same manner, and with different abiotic and biotic stresses and genes.

**Table 1:** Genes being measured, their functions, and primer sequences.

<table>
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<th>Gene function</th>
<th>Sequences</th>
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<td><em>rd22</em></td>
<td>(responds to dehydration)</td>
<td>Forward 5’ ATGGATCCTGCTTATGGCCG 3’&lt;br&gt;Reverse 5’ TCTTGCAGCAACAACCTCTG 3’</td>
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<tr>
<td><em>rab18</em></td>
<td>(responds to abscisic acid)</td>
<td>Forward 5’ AGAAGGGCATCATGGACAAG 3’&lt;br&gt;Reverse 5’ TCACACTGCACACACAGGTG 3’</td>
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<td><em>dreb2a</em></td>
<td>(dehydration response element binding)</td>
<td>Forward 5’ GTGCCTTCACAGTGCTTGAA 3’&lt;br&gt;Reverse 5’ AAGAGTTGAGGAAGCGCCAA 3’</td>
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<tr>
<td><em>ap2</em></td>
<td>(involved in chromatin segregation)</td>
<td>Forward 5’ GTGCCTTCACAGTGCTTGAA 3’&lt;br&gt;Reverse 5’ AAGAGTTGAGGAAGCGCCAA 3’</td>
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<td><em>ap1</em></td>
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### 2. Objectives

- Grow and apply drought and non-drought treatments to Alamo and Cave-In-Rock genotypes.
- Compare expression levels of the selected genes under drought stress between the two genotypes.

### 3. Materials and Methods

- **Plant Materials** - Alamo and Cave-In-Rock were grown from seed in a DSU greenhouse for approximately three months. Alamo is a lowland ecotype derived from Texas and Cave-In-Rock is an upland ecotype from Missouri.
- **Methods** - Field capacity was determined using a percolation method, and control plants received a full watering of 355mL when soil moisture fell below 20% VWC (volumetric water content), while drought plants received half (177mL) when soil moisture fell below 6%.
- Leaves were harvested and stored in -80°C until RNA isolation.
- RNA was isolated using Total RNA isolation kit by Machery-Nagel (Bethlehem, PA).
- cDNA was synthesized using cDNA synthesis kit by NEB Protoscript (Ipswich, MA).
- qPCR was conducted using 7500 Real-time PCR System and SYBR Green Master Mix by Applied Biosystems (Grand Island, NY).
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### 4. Results

**Figure 1:** Drought treated Alamo (left) adjacent to control Alamo (right).

**Figure 2:** Isolated RNA

**Figure 3:** PCR of cDNA

**Figure 4:** Amplification plot of qPCR reactions

**Figure 5:** Amplitization plot of qPCR reactions

### 5. Future Directions/Discussion

- Perform more biological replicates of the qPCR procedure.
- Cultivate and test different genotypes in the same manner, and with different abiotic and biotic stresses and genes.

**D. Aimar et all., “Drought effects on early developmental stages of Panicum virgatum L: cultivar differences” Biomass and Bioenergy, 2014**

### 6. References

We would like to thank the NEWBio program for funding this research (grant no: 2012-68005-19703), Delaware State University for hosting us, and everyone in the MGE lab for their support.

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